Synthesis of Quinolines from 3-Formylchromone

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A facile and versatile procedure for the synthesis of 3-(2hydroxybenzoyl)quinolines and 7*H*-chromeno[3,2-*c*]quinolin-7-ones was elaborated on the basis of TMSCI-mediated recyclization of 3-formylchromone with various anilines. Limitations and scope of this methodology were established, and a possible mechanism for the heterocyclizations was proposed.

The functionalized quinolines are attractive compounds for drug discovery since many of them have been shown to exhibit excellent biological activities.¹ Therefore, the development of facile methodologies for the synthesis of highly functionalized quinoline derivatives represents a challenge in medicinal chemistry.^{2,3} For example oxoindeno[1,2-b]quinolines have shown strong binding to DNA and efficient inhibition DNA topoisomerase 1 that were associated with their well pronounced anticancer activities.⁴ 3-Benzoylquinolines^{5,6} are inhibitors of HIV-1 replication^{7a} and cathepsin D^{7b} and possess antianaphylactic activity.^{7c} The Combes synthesis is one of the most

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efficient approaches to the quinolines. It occurs through condensation of 1,3-dicarbonyl compounds with primary anilines followed by acid-catalyzed ring closure of the intermediate Schiff bases.^{8–11} 3-Fomylchromone **1** is a latent 1,3-dialdehyde bearing a masked 2-hydroxybenzoyl fragment at the meso-position.^{12,13} Recently we have shown that the recyclization and [3 + 3] cyclocondensation of 3-formylchromones with CH-active compounds such as cyanoacetamides,^{14a} amino heterocycles,^{5a} and benzimidazoles^{14b} readily gives functionally diverse sub-

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JOC Note

SCHEME 1



SCHEME 2



stituted pyridines. The present study was undertaken in order to develop a facile method for the synthesis of 3-(2-hydroxy-benzoyl)quinolines through the coupling of 3-formylchromone 1 with anilines.

On the basis of our previous studies, ^{5a,14} we considered TMSCl as a potential promoter and water scavenger for the reaction of 3-formylchromone with anilines unsubstituted at the ortho-position that might result in various o-hydroxybenzoyl quinolines. The present research was undertaken in order to determine limitation and establish the scope of this reaction with various substituted anilines. The reaction of substituted anilines 2a-l with 3-formylchromone gave quinolines 3a-l in 35-87% yield. (Scheme 1). It seems likely that the reactivity of the orthoposition in substituted anilines is influenced by +M and sterical effect of the functional groups in meta- and para-positions. This indicates that on the first bimolecular step of the reaction anilines 2 react as C-nucleophiles. The reactions of the ortho-substituted anilines with formylchromone resulted in multicomponent mixtures, which according to LCMS contain less than 20% of the corresponding quinolines 3.

Surprisingly, the reactions of anilines $4\mathbf{a}-\mathbf{i}$ with 3-formylchromone (TMSCl, DMF) led to 7*H*-chromeno[3,2-*c*]quinolin-7-ones¹⁵ $5\mathbf{a}-\mathbf{i}$ (Scheme 2) in 39–63% yield. No quinolines of type **3** were detected in the reaction mixtures by LCMS. The fact that the substituents in molecules **4** withdraw electrons from the *ortho*-positions to the amino group or increase the electron density on the nitrogen atom indicate that anilines **4** react as N-nucleophiles. The reaction of 3-chloroaniline with 3-formylchromone gave a multicomponent mixture containing the corresponding compounds **3** (33%) and **5** (14%) and three unidentified products. 3,4-Dichloroaniline **4j** whose C-nucleophilicity is decreased by the presence of a chlorine atom at the *meta*-position to the amino group reacted with **1** to give chromenoquinolin-7-one **5j** in 51% yield.

TMSCl can activate 3-formylchromone 1 through addition to aldehyde or ketone C=O bonds to give intermediates **6a** and **6b**, respectively¹⁶ (Scheme 3).

Apparently intermediate **6a** is more electrophilic and more reactive than resonance-stabilized *O*-trimethylsilyl-3-formylchromone chloride **6b**. Scheme 4 outlines possible mechanisms for the formation of quinolines **3** and **6** starting from activated 3-formylchromone **6a**. Amines **2** are likely to react with **6a** as C-nucleophiles to give intermediate **7**, which after activation of the keto group and double bond of chromone undergoes intramolecular N-arylation to give intermediate **8**. Elimination of HMDS from **8** can result in N-silylated quinolyl chromone **9**, which can be transformed into trimethylsiloxybenzoyl quino-

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SCHEME 4



line 10 through intramolecular migration of the trimethylsilyl group from the nitrogen to oxygen atom and breaking the C-O bond.

The reaction of N-nucleophilic amine 4 with intermediate 6a seems likely to give intermediate 11, which after activation of the keto group and the adjacent double bond of chromone undergoes intramolecular cyclization through electrophilic substitution at the ortho-position to the aniline nitrogen atom. The intramolecular elimination of HMDS is likely to result in the silvlated dihydroquinoline 13 intermediate, which can be transformed into compound 5 through the loss of hydride and hydrolysis. It seems plausible that the long distance between the silicon and oxygen atom at position 4 makes impossible the intramolecular migration of the trimethylsilyl group resulting in the opening of the pyrone ring. After quenching of the reaction mixture, dihydroquinoline 14 seems likely to disproportionate into final product 5 and tetrahydroquinoline 15. Accordingly, the LCMS studies of the reaction mixtures revealed the presence of tetrahydroquinolines 15 as a byproduct of quinolines 5. Unfortunately the preparative HPLC failed to isolate individual compounds 15.

Similar considerations indicate that the reaction of intermediate **6b** with C-nucleophilic anilines **2** and N-nucleophilic anilines **4** should give quinolines **5** and **3**, respectively (see Scheme 1 in Supporting Information). This provides an additional argument in favor of the mechanism shown in Scheme 4.

The composition and structure of all the compounds obtained were determined by LC/MS, elemental analysis, and ¹H and ¹³C NMR and IR spectroscopy. The signals in the ¹H and ¹³C NMR spectra were assigned on the basis of 2D NMR techniques (COSY, NOESY, HMBC, HMQC) of representative compounds. The typical ¹H NMR spectrum of quinolines **3** contains a characteristic set of signals for the protons of *ortho*-acylated phenol ring, and two doublets (⁴*J*_{HH} \approx 2.0 Hz) or broadened singlets for the protons of the pyridine fragment. The signal of the OH group appears in the range of 10.4–10.6 ppm (DMSO- d_6) and is deshielded as a result of the formation of the stable intramolecular bond to the adjacent carbonyl group. The typical 13 C NMR spectrum of quinolines **3** contains the signal of a carbonyl carbon atom at $\delta \approx 193.1-196.6$ ppm and a characteristic set of signals for the carbons of ortho-acylated phenol ring and for the carbons of the pyridine fragment. In the typical IR spectra of quinolines 3, a wide absorption band at $3650-3300 \text{ cm}^{-1}$ corresponding to valence vibrations of the hydroxyl group, and an intensive peak at 1622–1639 cm⁻¹ corresponding to valence vibrations of carbonyl group is present. The ¹H NMR spectra of compounds 5 contains a set of signals for the ortho-substituted aryl ring of the chromone residue and a singlet for the methine proton of the quinoline fragment $(9.3-9.6 \text{ ppm in DMSO-} d_6 \text{ or } 9.9-10.2 \text{ in CF}_3 \text{COOD})$, whereas no signal was detected for the phenolic OH group. The ¹³C NMR spectra of compounds 5 contains the signal of carbonyl carbon atom at $\delta \approx 175.5 - 175.8$ ppm (DMSO-d₆) or $\delta \approx 175.8 - 185.5$ ppm (CF₃COOD), a set of signals for the *ortho*-substituted aryl ring of the chromone residue, and a signal for the C-2 of the quinoline fragment (145.5-148.7 ppm in DMSO-d₆ or 145.2-148.1 in CF₃COOD).

The structure of compound **5k** was unambiguously determined by single crystal X-ray analysis¹⁷ (see Figure 1, X-ray crystal structure determination, in Supporting Information). In the crystalline state plane molecules of **5k** are involved in stacking interactions to give infinite columns in the crystal.

In conclusion, we have elaborated efficient synthetic procedures for the preparation of functionalized quinolines from 3-formylchromone and substituted anilines using TMSCI as a promoter and water scavenger. The reaction of 3-formylchromone with C-nucleophilic anilines **2** or N-nucleophilic anilines **4** leads to 3-(2-hydroxybenzoyl)quinolines and 7*H*chromeno[3,2-*c*]quinolin-7-ones, respectively. Apparently the

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developed procedure can be applied for the synthesis of diverse sets of functional drug-like quinolines.

Experimental Section

Preparation of 3-(2-Hydroxyphenyl)quinolines 3a-1 and 7H-Chromeno[3,2-c]quinolin-7-ones 5a-o from Anilines 2a-1 or 4a-o and 3-formylchromone 1. General Procedure. Anilines 2a-1 or 4a-o (2 mmol) and 3-formylchromone 1 (348 mg, 2 mmol) were placed in a 15-mL pressure tube and dissolved in DMF (2-4 mL). Chlorotrimethylsilane (652 mg, 3 mmol) was added dropwise to the solution. The tube was thoroughly sealed and heated on a water bath (100 °C) for 12-24 h. After cooling the flask was opened (*Caution! Excessive pressure inside*), and the reaction mixture was poured into water (15 mL) and allowed to stand at 20 °C in an ultrasonic bath for 1 h. The precipitate formed was filtered and washed with a small amount of *i*-PrOH. Recrystallization from an appropriate solvent yielded targeted compounds 3a-1 and 5a-o. Compounds 3i and 5g were purified by preparative HPLC.

8-(2-Hydroxybenzoyl)-2-methyl-2H-[1,4]oxazino[2,3-g]quinolin-3(4H)-one (3c). Yield 87%; mp 272-273 °C (EtOH-DMF). ¹H NMR (500 MHz, DMSO- d_6): δ 1.50 (d, ³ $J_{H,H}$ = 6.6 Hz, 3H, CH<u>CH₃</u>), 4.92 (q, ${}^{3}J_{H,H} = 6.6$ Hz, 1H, <u>CH</u>CH₃), 6.98 (t, ${}^{3}J_{H,H} =$ 8.1 Hz, 1H, 5-H_{Ar}), 7.02 (d, ${}^{3}J_{H,H} = 8.1$ Hz, 1H, 3-H_{Ar}), 7.40–7.51 (m, 3H, 4,6-H_{Ar}, 5-H_{Qn}), 7.53 (s, 1H, 8-H_{Qn}), 8.53 (s, 1H, 4-H_{Qn}), 8.93 (s, 1H, 2-H_{On}), 10.38 (br. s, 1H, OH), 11.27 (br. s, 1H, NH). ¹³C NMR (125 MHz, DMSO- d_6): δ 17.0 (qd, ¹ J_{CH} = 129.2 Hz, ${}^{2}J_{\text{CH}} = 3.7$ Hz, CH₃), 73.6 (dq, ${}^{1}J_{\text{CH}} = 151.2$ Hz, ${}^{2}J_{\text{CH}} = 4.2$ Hz, <u>C</u>HCH₃), 112.6 (dd, ${}^{1}J_{CH} = 164.5$ Hz, ${}^{3}J_{CH} = 5.0$ Hz, 5-C_{Qn}), 113.6 (d, ${}^{1}J_{CH} = 164.1$ Hz, 8-C_{Qn}), 117.6 (dd, ${}^{1}J_{CH} = 161.8$ Hz, ${}^{2}J_{CH} =$ 7.4 Hz, 3-C_{Ar}), 119.8 (dd, ${}^{1}J_{CH} = 129.2$ Hz, ${}^{2}J_{CH} = 7.6$ Hz, 5-C_{Ar}), 123.3 (d, ${}^{2}J_{CH} = 5.0$ Hz, 3-C_{Qn}), 124.7 (m, 1-C_{Ar}), 129.7 (d, ${}^{2}J_{CH}$ = 7.3 Hz, 4a-C_{Qn}), 130.0 (d, ${}^{2}J_{CH}$ = 6.8 Hz, 7-C_{Qn}), 131.4 (ddd, ${}^{1}J_{CH} = 160.3 \text{ Hz}, {}^{2}J_{CH} = 7.4 \text{ Hz}, {}^{3}J_{CH} = 1.8 \text{ Hz}, 6 \cdot C_{Ar}$, 134.4 (dd, ${}^{1}J_{CH} = 160.4 \text{ Hz}, {}^{2}J_{CH} = 8.7 \text{ Hz}, 4 \cdot C_{Ar}$), 137.2 (dt, ${}^{1}J_{CH} = 165.0$ Hz, ${}^{3}J_{CH} = 5.0$ Hz, 4-C_{Qn}), 147.1 (m, 6-C_{Qn}), 148.4 (m, 8a-C_{Qn}), 148.8 (dd, ${}^{1}J_{CH} = 181.9$ Hz, ${}^{3}J_{CH} = 6.0$ Hz, $2\text{-}C_{Qn}$), 158.0 (m, 2- C_{Ar}), 167.5 (t, ${}^{2}J_{CH} = 3.7$ Hz, CONH), 196.4 (m, C=O). IR (KBr), ν_{max} (cm⁻¹): 3650–3300 (br, OH, NH), 3049, 2995, 2951, 1704 (C=O_{amide}), 1624 (C=O), 1593, 1512, 1462, 1377, 1348, 1304, 1248, 1219, 1159, 1097, 1041, 920, 759. APSI MS: M⁺ + 1 = 335. Anal. Calcd for C₁₉H₁₄N₂O₄: C, 68.26; H, 4.22; N, 8.38. Found: C, 68.42; H, 4.09; N, 8.31.

3-Methyl-2-(2-oxopyrrolidin-1-yl)-7H-chromeno[3,2-c]quinolin-7-one (5k). Yield 67%; mp 281-282 °C (MeCN). ¹H NMR (500 MHz, DMSO- d_6): δ 2.24 (quintet, ${}^{3}J_{H,H} = 6.9$ Hz, 2H, NCH_2CH_2), 2.42 (s, 3H, CH₃), 2.43 (t, ${}^{3}J_{H,H} = 6.9$ Hz, 2H, COCH₂), 3.90 (t, ${}^{3}J_{H,H} = 6.9$ Hz, 2H, NCH₂), 7.57 (t, ${}^{3}J_{H,H} = 8.2$ Hz, 1H, 6-H_{Chr}), 7.89 (d, ${}^{3}J_{H,H} = 8.2$ Hz, 1H, 8-H_{Chr}), 7.96 (t, ${}^{3}J_{H,H} = 8.2$ Hz, 1H, 7-H_{Chr}), 8.04 (s, 1H, 8-H_{Qn}), 8.23 (d, ${}^{3}J_{H,H} = 8.2$ Hz, 1H, $5\text{-}H_{Chr}),\ 8.46\ (s,\ 1H,\ 5\text{-}H_{Qn}),\ 9.37\ (s,\ 1H,\ 2\text{-}H_{Qn}).\ ^{13}C\ NMR\ (125)$ MHz, DMSO-*d*₆): δ 18.9 (CH₃), 19.4 (NCH₂CH₂), 31.3 (COCH₂), 50.9 (NCH₂), 112.6 (3-C_{Chr}), 117.2 (4a-C_{Chr}), 119.1 (8-C_{Chr}), 120.8 (8-C_{Qn}), 123.4 (4a-C_{Qn}), 126.1 (6-C_{Chr}), 126.3 (5-C_{Chr}), 131.0 (5- C_{Qn}), 136.2 (7- C_{Chr}), 139.3 (6- C_{Qn}), 142.5 (7- C_{Qn}), 148.7 (2- C_{Qn}), 149.2 (8a-CQn), 155.8 (2-CChr), 158.6 (8a-CChr), 174.6 (COCH2), 175.6 (C=O). IR (KBr), v_{max} (cm⁻¹): 3070, 3041, 2962, 2920, 1682 (C=O_{amide}), 1662 (C=O), 1628, 1614, 1495, 1470, 1446, 1414, 1362, 1294, 1223, 899, 827, 756. APSI MS: M⁺ + 1 = 345. Anal. Calcd for C₂₁H₁₆N₂O₃: C, 73.24; H, 4.68; N, 8.13. Found: C, 73.08; H, 4.82; N, 8.21.

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Supporting Information Available: Details of the experimental procedures, spectroscopic data of the products, and X-ray data for the compound **5k** in CIF format. This material is available free of charge via the Internet at http://pubs.acs.org.

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